



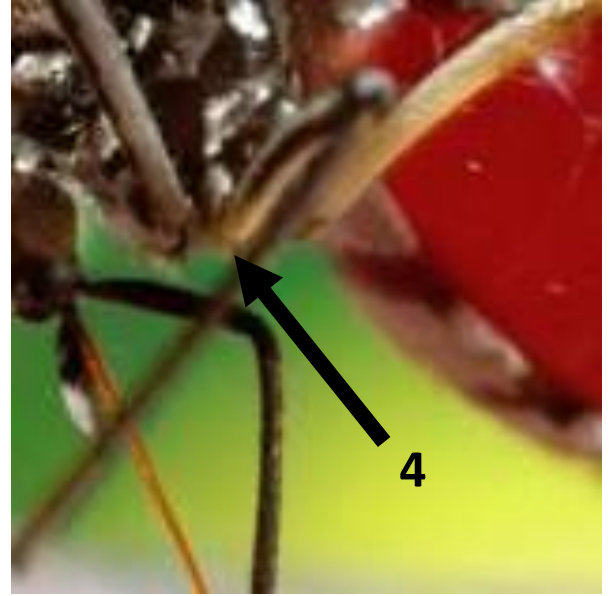
Shining light on invasive and  
non-native mosquito species

# Invasive and Non-Native Adult Female Mosquitoes Found in Southeastern United States & Emerging Invasive Species Not Yet Established in Continental US

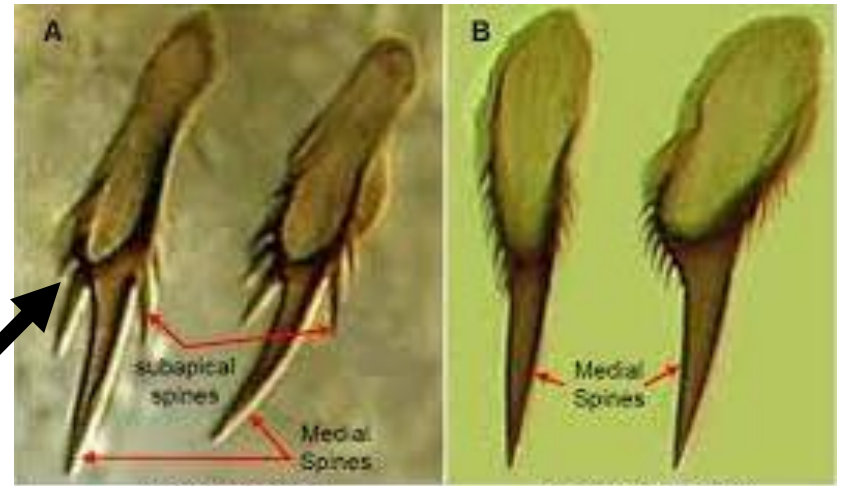
Mosquito BEACONS Working Group

Contributors: Benjamin Allen, Bryan V. Giordano, Michael Riles, Ana L. Romera-Weaver, Yoosook Lee

# *Aedes aegypti*



1. Lyre shaped patch of white scales on Dorsal Thorax
2. White scales on Clypeus
3. Solid white abdominal sternites
4. White Band on dorsal middle femur
5. Larval comb scales with prominent sub-apical spines



*Aedes aegypti*

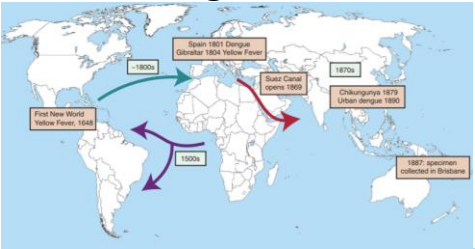
*Aedes albopictus*



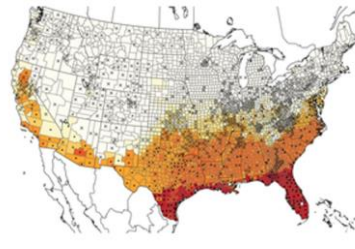
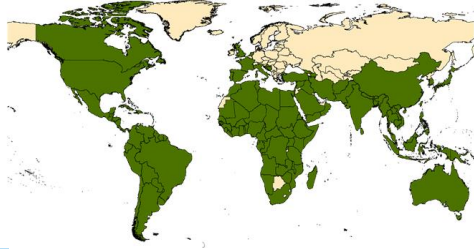
Mosquito BEACONS  
Working Group

# *Aedes aegypti*

## Native Range



## Invasive Range



## Collection Methods



- For larvae: turkey baster and dipping cups
- For adults: BG sentinel traps, BG Lyre baited CDC traps, aspirators, sweep nets

## Host Biting Preference

- Most active for roughly two hours after sunrise and several hours before sunset
- It can bite at night in well-lit areas.
- have a strong preference for human hosts
- occasionally feed on dogs, cats, horses and other domestic animals.

## Pathogen Transmission

- Transmit yellow fever virus, dengue virus, chikungunya virus, and Zika virus.
- potential vector of Venezuelan Equine Encephalitis virus and West Nile virus.

## Larval Habitat

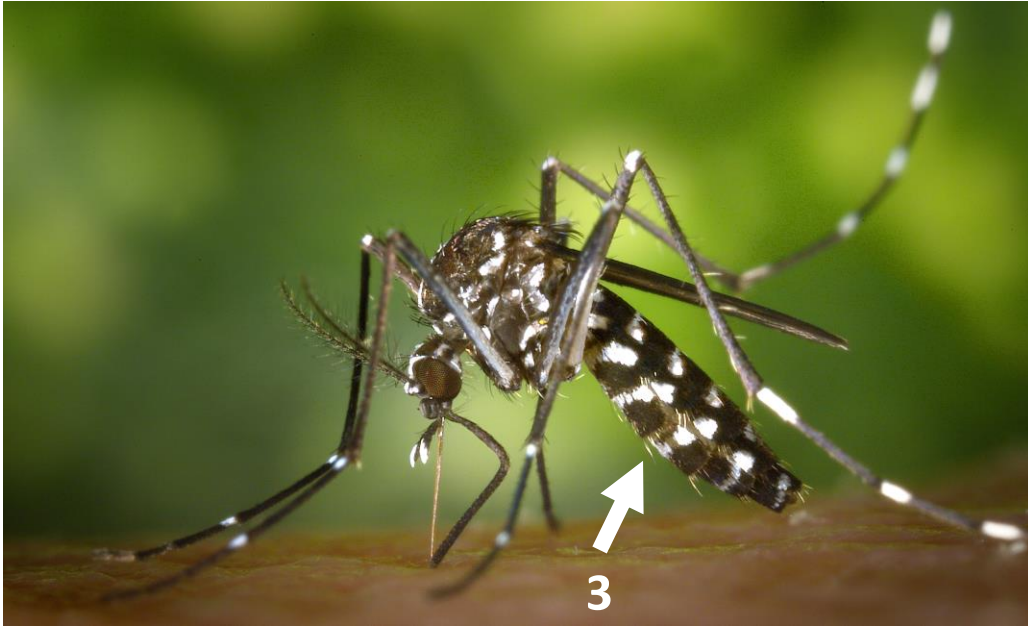


- Rotten tree stumps, tree holes, water-holding plants
- Vases, bird baths, gutters, tires

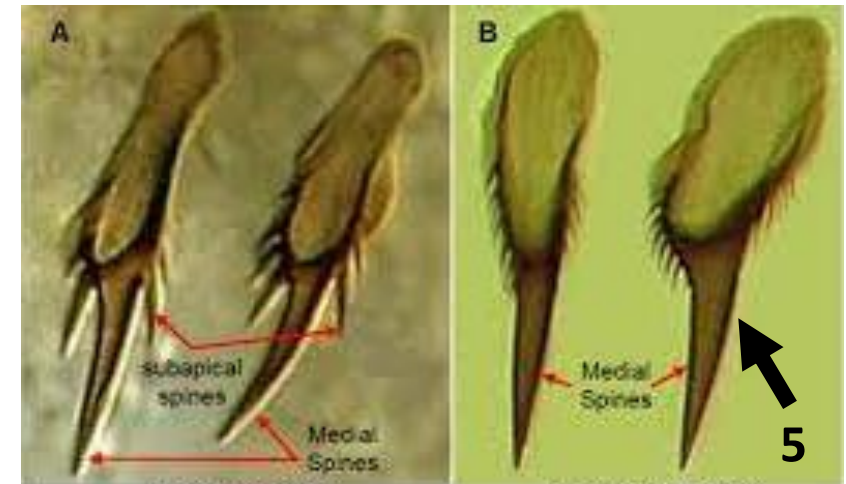
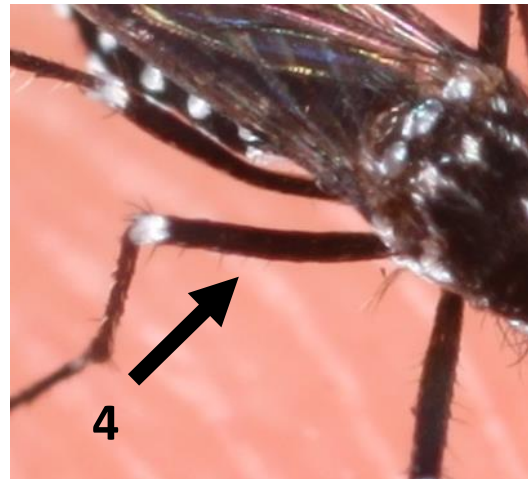
## Dispersal Strategies

- Short flight range (100–500 meters < 0.3 miles)
- Desiccation tolerant eggs
- Spread by human transport of containers

# *Aedes albopictus*



1. White stripe down dorsal thorax
2. Clypeus entirely dark, secondary differentiation *from Ae aegypti*
3. Abdominal sternites are white-banded
4. No white band on middle dorsal femur
5. Comb scales have only the apical spine



*Aedes aegypti*

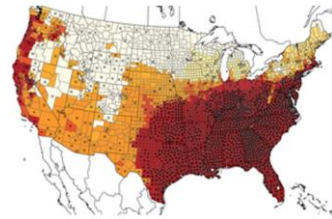
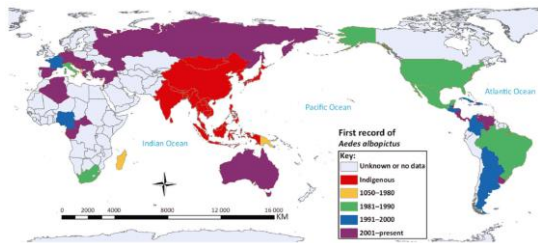
*Aedes albopictus*



Mosquito BEACONS  
Working Group

# *Aedes albopictus*

## Native (Red) and Invasive (other colors) Range



## Larval Habitat



- Various artificial (manmade) and natural containers
- Used tires, discarded plastic containers, tree holes

## Dispersal Strategies

- Egg desiccation tolerance
- Ornamental plant trade and used tire trade (internationally and locally)
- Even follow people into cars!

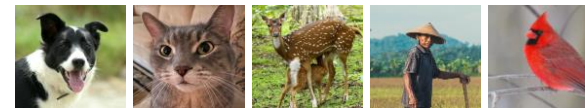
## Collection Methods



- Larval surveillance highly productive
- BG-Sentinel traps BG-Lure can be combined with CO<sub>2</sub> (dry ice or compressed gas) to increase yield of collections
- Not commonly collected with CDC light traps unless baited with BG Lure

## Host Biting Preference

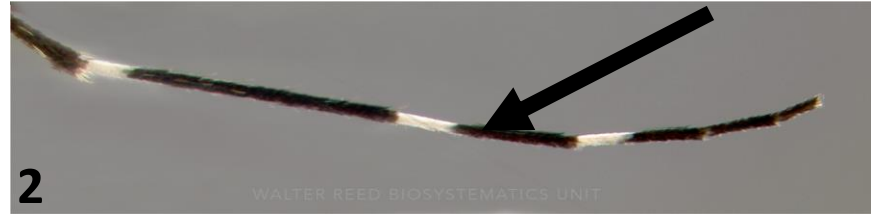
- Early morning and late afternoon
- Bite a variety of mammals (Humans, dogs, cats, ruminants)
- Opportunistic feeders and will occasionally bite birds



## Pathogen Transmission

- Transmit yellow fever virus, dengue virus, chikungunya virus, and Zika virus.
- potential vector of West Nile virus.

# *Aedes japonicus*



- 5. Siphon pecten spines >15 with two pronounced larger spines outside row
- 6. Seta 5,6-C >5 branches set in linear orientation
- 7. Anal Saddle conspicuously spiculated

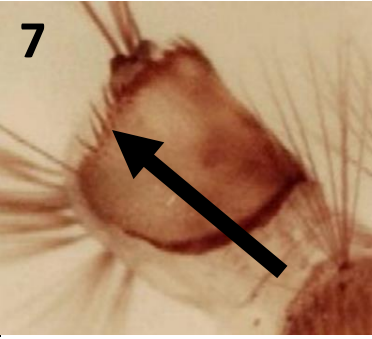
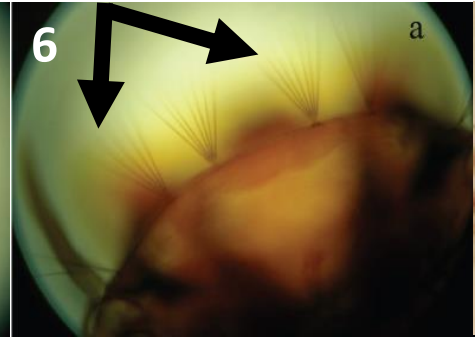
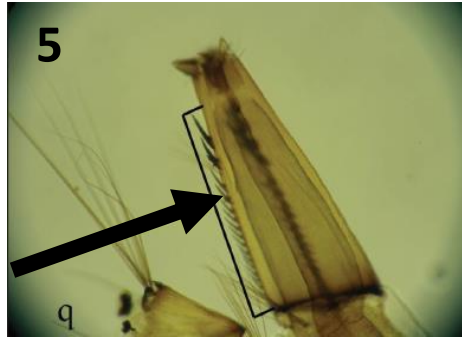
1. Medium to large mosquito. Banded legs, golden, dark-brown and pale scales throughout. Proboscis dark scaled not banded

2. Tarsi Ta-III 1–3 with broad pale basal bands; Ta-III5 all dark

3. Femoral character; pale scaled not touching insertion of thorax; origin at middle of femur

4. Scutum with golden stripes; distinctive lyre-shaped stripes, and two sub-median and a median stripe.

Scutellar lobes with long, narrow scales



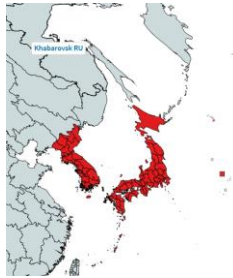
Larvae Photo Credit: George O'Meara UF/FMEL  
Adult Photo Credit: WRBU; Lyle J. Buss UF/FMEL



Mosquito BEACONS Working Group

# Aedes japonicus

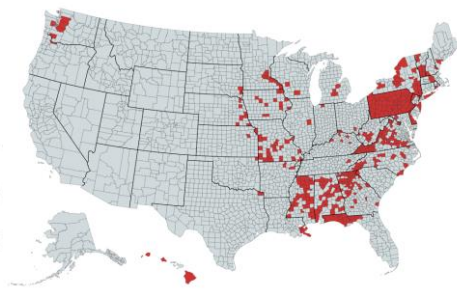
## Native Range



## Invasive Range



## Current Expansion 2022



- Slower expansion than *Aedes albopictus*
- Introduced into North America & Europe in mid-late 1990's
- Florida, Oklahoma, Kansas, Louisiana, Texas, & Nebraska 2014-2021
- Need better collection records

## Larval Habitat



- Rock pools in native range
- Oviposit on the walls of natural and artificial containers
- Examples: Tree holes, tires, containers made of concrete, stone, plastic, or metal

## Dispersal Strategies

- Egg desiccation tolerance
- Commercial transport (tires, botanicals, construction)
- Ports of entry (seaports, airports, bio-security)

## Collection Methods



- Larval surveillance
- Not readily trapped with CDC light traps + CO<sub>2</sub>
- octenol-based lures more effective
- Gravid and oviposition traps are ideal
- BG-Sentinel traps
- Aspirations work well

## Host Biting Preference

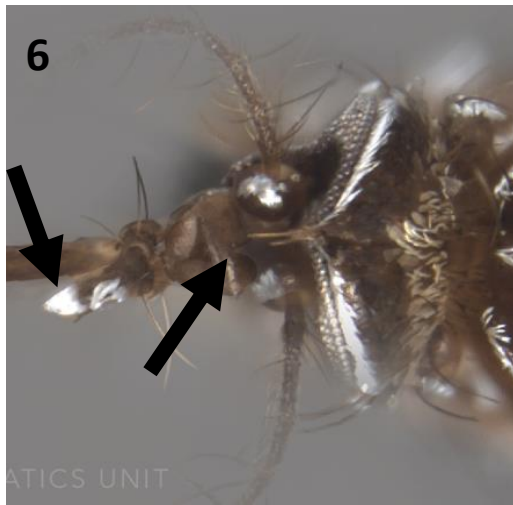
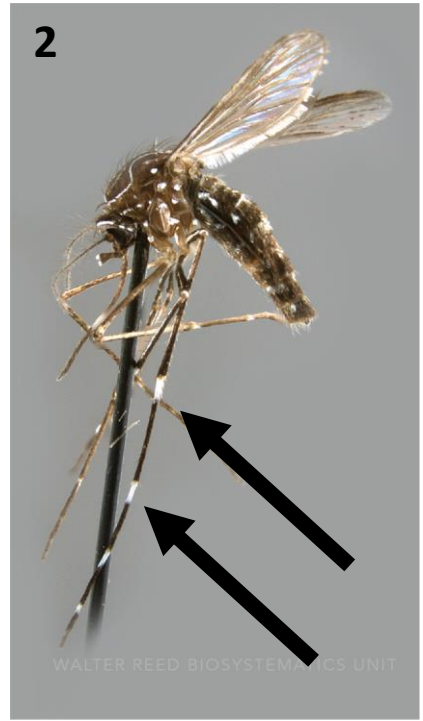
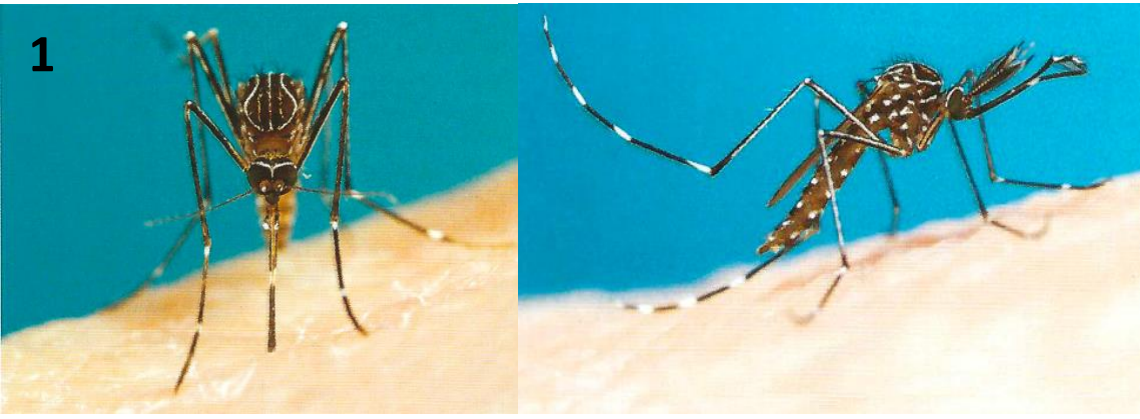
- Feeding peaks early morning and late afternoon
- Leave shady or wooded areas to feed
- Opportunistic, wide-range of host species



## Pathogen Transmission

- Chikungunya virus (CHIKV)
- Cache Valley virus (CVV)
- Dengue virus (DENV)
- Eastern Equine Encephalitis virus (EEEV)
- Japanese Encephalitis virus (JBEV)
- Lake Victoria cormorant virus (LVC)
- Usutu virus (USUV)
- LaCrosse Virus (LACV)
- Jamestown Canyon virus
- Orungo virus (ORUV)
- Rift Valley fever virus (RVFV)
- St. Louis Encephalitis virus (SLEV)
- Western Equine Encephalitis virus (WEEV)
- West Nile virus (WNV)
- Zika (ZIKV)

# *Aedes notoscriptus*



1. Female & Male Small to mid-sized species; Integument can vary in color from reddish brown to dark brown to black

2. Fe-II and Ti-III with anterior lines of pale scales along nearly whole length  
Hind tarsi with wide bands bright silvery whitish scales

3. Proboscis dark-scaled with median pale ring  
Wings scales all dark dorsally

4. Scutum with lyre-shaped lateral silvery lines

- Narrow median silvery lines and anterior sublateral golden lines of scales
- Postpronotum with small distinct lower patch of silvery scales

5. Small patches of silvery scales throughout thoracic area

6. Pedicel not scaled exteriorly only interiorly. Palps tipped with silvery scales

7. Terminal segments Larvae: Comb scales in irregular triangular patch; siphon not pilose; siphonal acus large and broadly joined to siphon; seta 1-S short, <0.5 x length of siphon





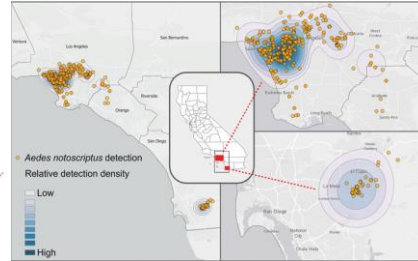
Mosquito BEACONS  
Working Group

# *Aedes notoscriptus*

## Native Range



## Invasive Range



- Established in New Zealand around 1920
- Migrated north into South-east Asia & South Pacific Islands
- Established in 3 counties in southern California 2014-present time
- Cryptic in morphology regionally suggesting a diverse phenotypic morphologic complex

## Larval Habitat



- Wide range of artificial & natural containers
- Bamboo stumps, leaf axils, rock pools, palm fronds
- Containers made of wood, concrete, plastic and metal

## Dispersal Strategies

- Container breeders that use diapausing eggs
- Spread by humans along shipping routes

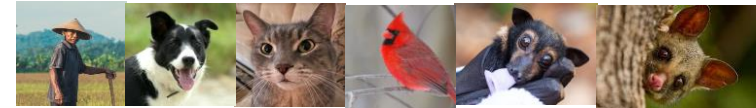
## Collection Methods



- CDC trap with CO<sub>2</sub> and Octenol or BG Lure
- BG Sentinel
- Autocidal Gravid Traps
- Gravid Traps

## Host Biting Preference

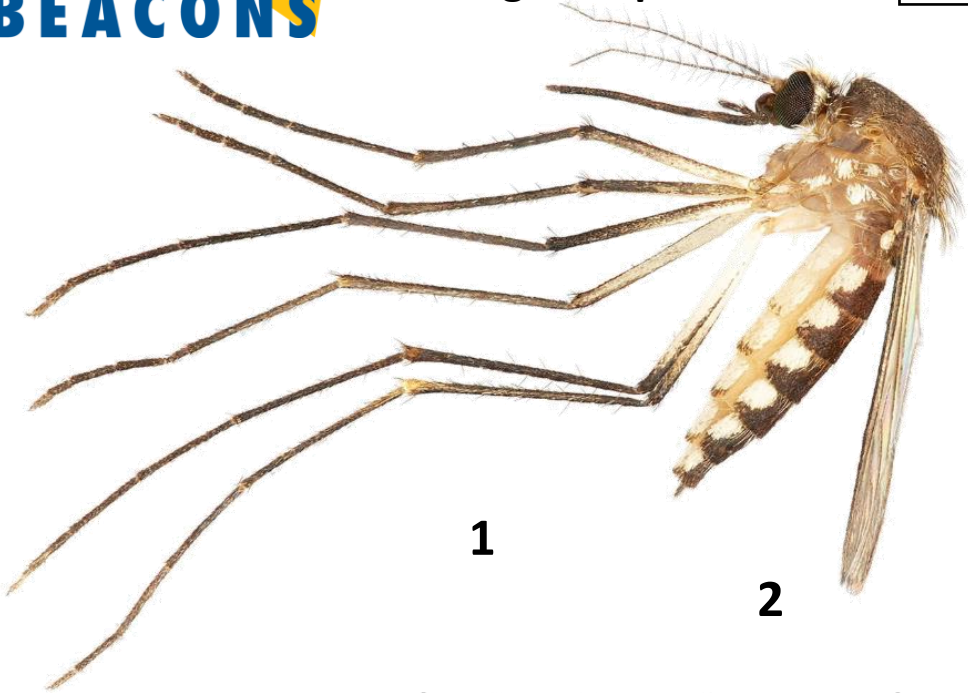
- Humans, domestic pets, birds, marsupials, bats



## Pathogen Transmission

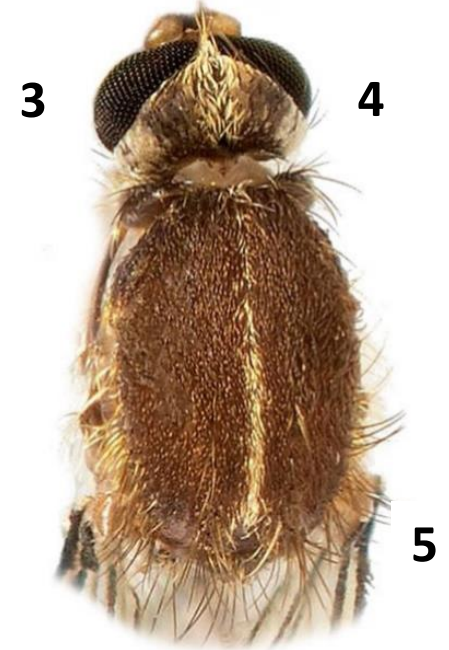
- Barmah Forest Virus
- Chikungunya virus (CHIKV)
- Dengue virus (DENV)
- Ross River Virus
- Zika (ZIKV)
- *Dirofilaria* (dog heartworm parasite)

# *Aedes pertinax*



1

2



3

4

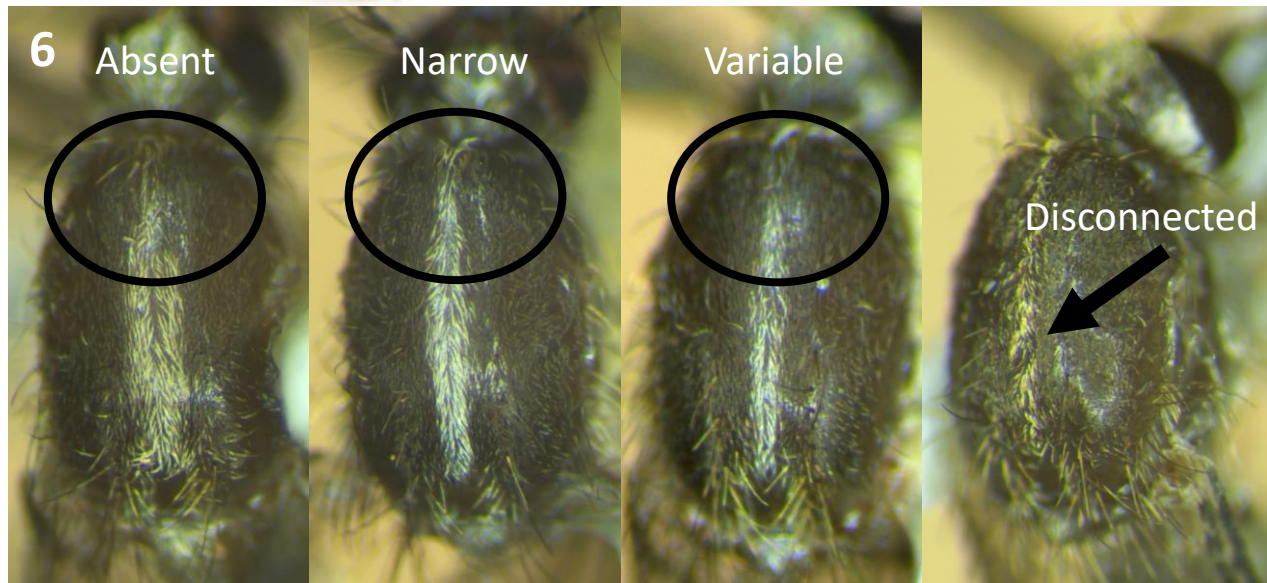
5



*Aedes atlanticus*

- 1. Hindtibia without basal and apical dark-scaled bands
- 2. Unbanded abdomen
- 3. Occiput clothed medially with white or yellow scales,
- 4. White to yellow scales extended on interocular space and along orbital line

- 5. Longitudinal stripe extending from anterior margin to full length of scutum
- 6. Scutum with narrow acrostichal line of whitish scales, absence of pale scales on anterior end, extremely thin anterior stripe, or disconnected exceptionally narrow stripe



6

Absent

Narrow

Variable

Disconnected

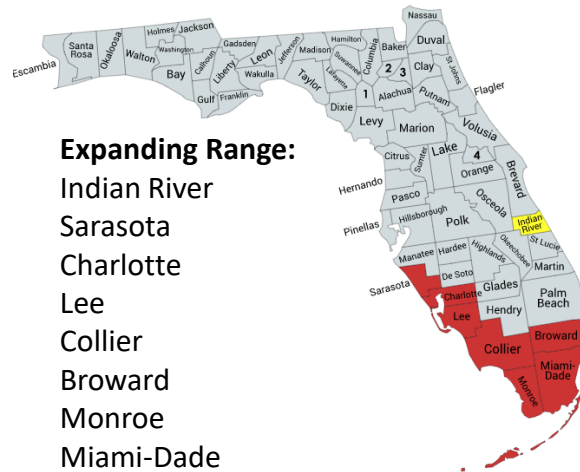
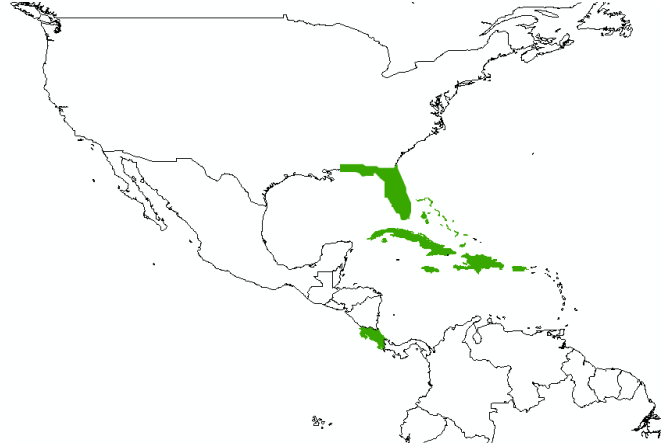


Mosquito BEACONS Working Group

# Aedes pertinax

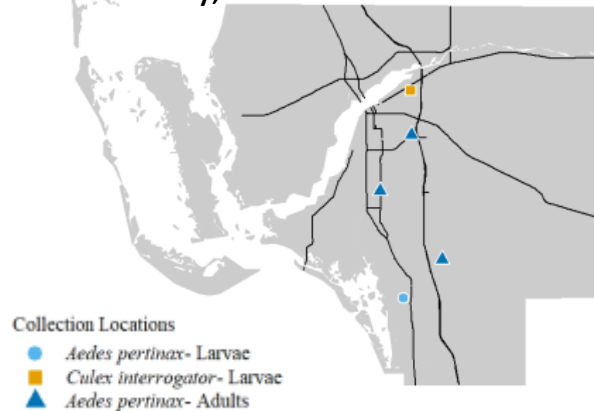
## Native and Invasive Range

- Native to South America and Caribbean
  - Actual distribution widely unknown
- Invasive in Florida



Lee County, FL 2021

Charlotte County, FL 2022



## Larval Habitat

- More research is needed in this area
- Coastal flooded woodlands
- Temporary and permanent grasslands / woodlots prone to flooding
- In Florida, *Ae. pertinax* was found in freshwater sites
  - E.g., nontidal, rain-filled depression



## Dispersal Strategies

- More research is needed in this area
- No flight distance is recorded for this species
- Hypothesis: Major storm winds driving dispersal



## Collection Methods

- Can be very abundant in CDC and BGS collections
- Larval collections from floodwater (freshwater)



## Host Biting Preference

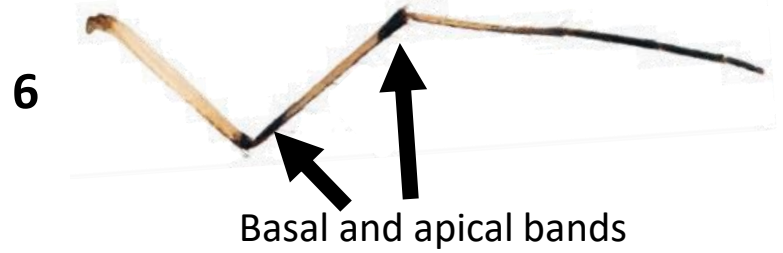
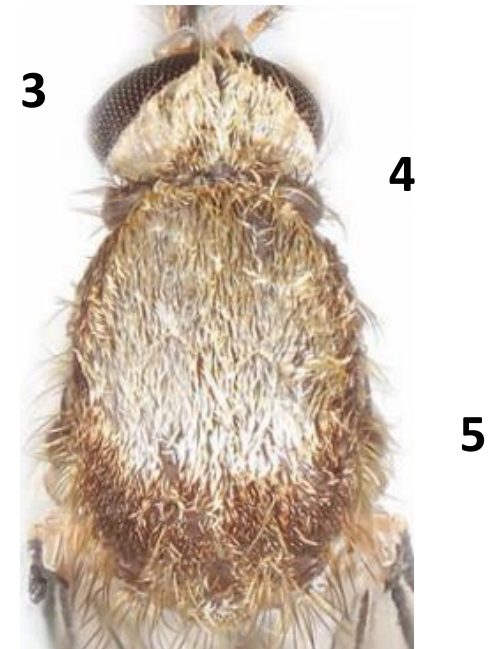
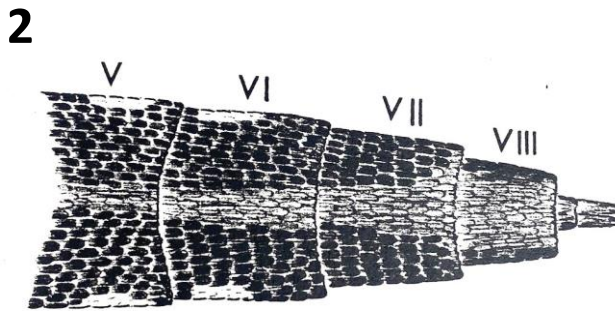
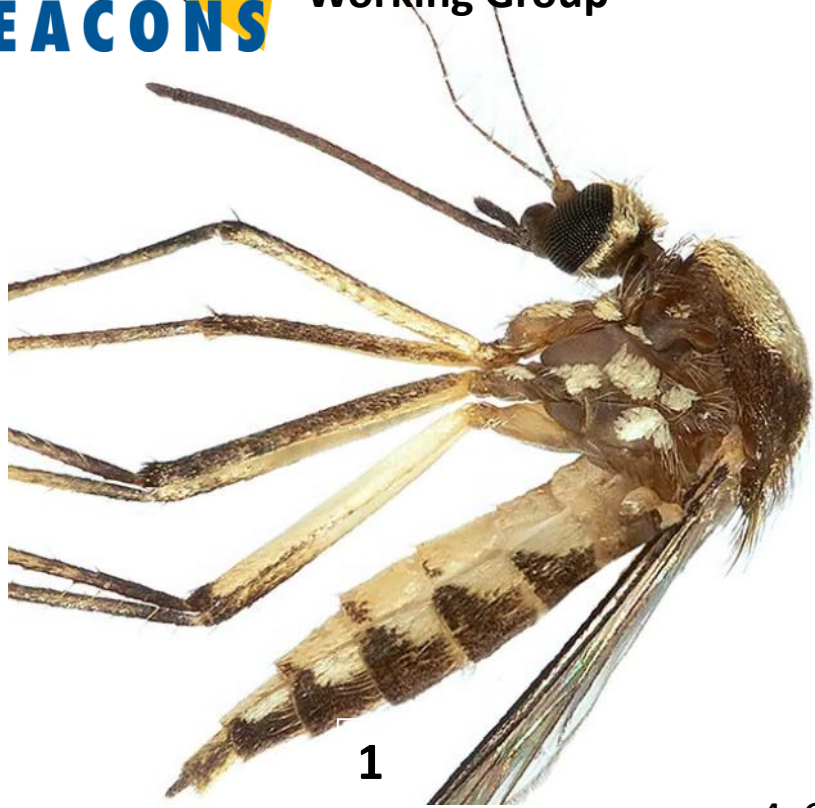
- More research is needed in this area



## Pathogen Transmission

- Nothing none to date

# *Aedes scapularis*



- 1. Unbanded abdomen
- 2. Abdominal terga VI-VIII with light colored scales medially
- 3. White to yellow scales extended on interocular space and along orbital line

- 4. Occiput clothed medially with white or yellow scales
- 5. Scutum with median longitudinal stripe extending from anterior margin to ~2/3 length of scutum
- 6. Hindtibia with basal and apical dark-scaled bands

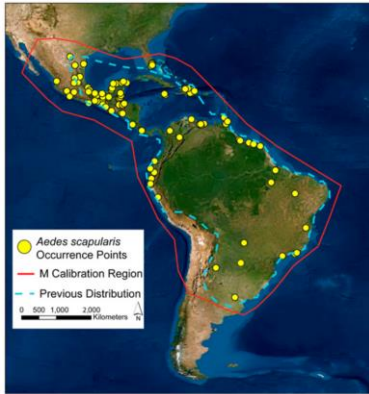
Basal and apical bands



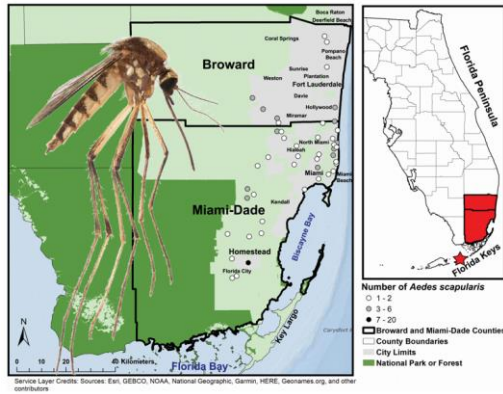
Mosquito BEACONS Working Group

# Aedes scapularis

Native Range



Invasive Range (as of 2021)



Expanding range in south Florida

- Monroe County (Florida Keys)
- Miami-Dade County
- Broward County
- Collier County

Collection Methods



- Rough equivalency in catch between CO<sub>2</sub>-Baited CDC traps, and BGS Traps

Larval Habitat



- Temporary and semi-permanent freshwater such as swamp margins, crab holes, overflowing waterways, and rain pools.
- Lays eggs in soil prone to flooding.
- Sometimes containers.

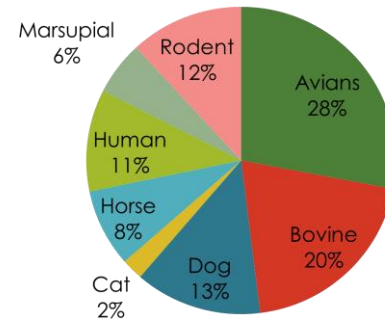
Dispersal Strategies

- Dispersal has been hypothesized to occur by aircraft, movement of plants and soil where eggs and larvae may be present



Host Biting Preference

Aedes scapularis Feeding Preference in Brazil

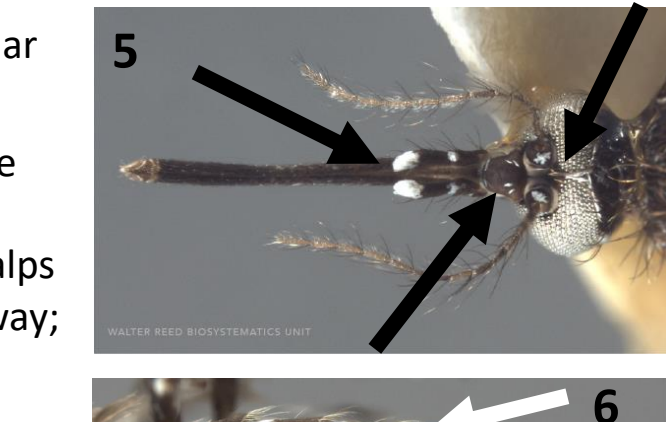
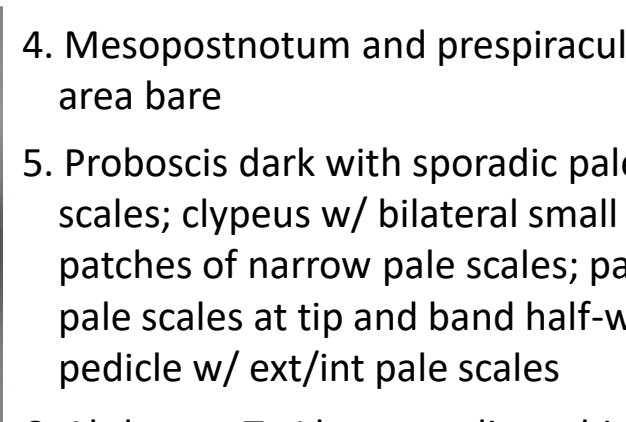
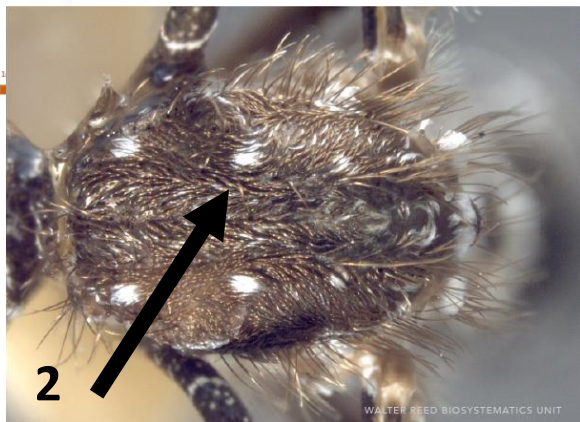
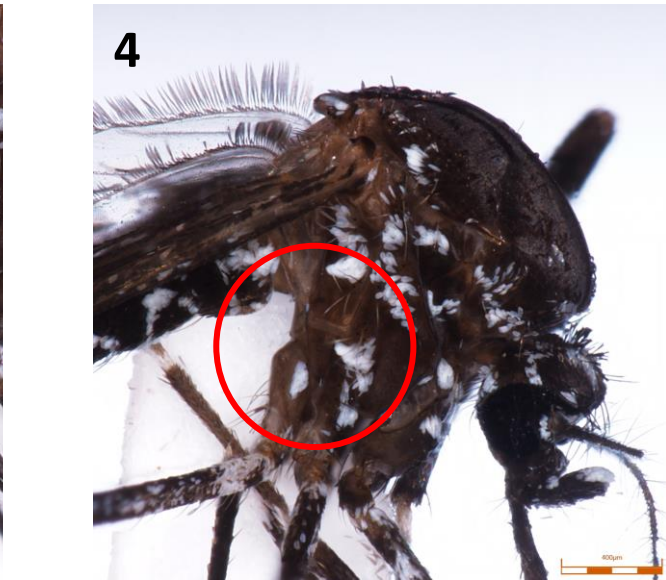
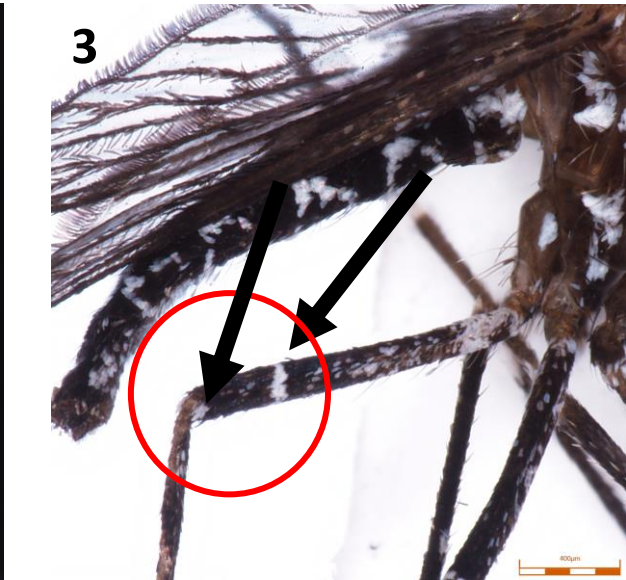


- Feeds in the middle of the night, on a wild variety of mostly-mammal hosts, including humans

Pathogen Transmission

- Melao virus (MELV)
- Oropouche virus (OROV)
- St. Louis Encephalitis virus (SLEV)
- Venezuelan Equine Encephalitis virus (VEEV)
- Yellow Fever virus (YFV)
- *Dirofilaria immitis* (Dog heartworm)

# *Aedes vittatus*



1. Small to medium dark scaled species with pale scales throughout

2. Scutum: 3 pairs of distinct, small, white spots of narrow scales on anterior two-thirds of scutum

3. Tibia dark, with sub-basal white spot and white band approximately level with basal third of Ti-I, Ti-II, and at mid-point of Ti-III

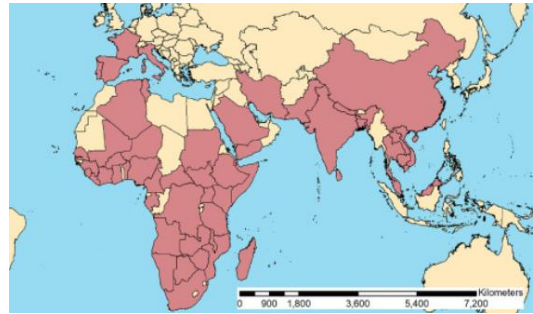
4. Mesopostnotum and prespiracular area bare

5. Proboscis dark with sporadic pale scales; clypeus w/ bilateral small patches of narrow pale scales; palps pale scales at tip and band half-way; pedicle w/ ext/int pale scales

6. Abdomen Te-I large median white spot; basal white bands w/ lateral curved white markings: disparate from bands

# *Aedes vittatus*

## Native Range



## Invasive Range (as of 2021)



## Larval Habitat



Variety of natural and artificial containers:

- rock pools
- tree holes
- hoofprints
- domestic containers

## Dispersal Strategies

- Egg desiccation tolerance
- Caribbean introductions are speculated to have occurred via shipping container transport to and between islands

## Collection Methods



- Like other invasive *Aedes* they are not commonly collected using CDC light traps
- Surveying potential production sites (artificial and manmade containers) is necessary
- More surveillance data in invasive range (Caribbean) is needed to predict production sites in North America

## Host Biting Preference

- Opportunistic feeders:
  - Non-Human Primates
  - Domestic Animals
  - Rodents
  - Humans
  - Bats
  - Birds

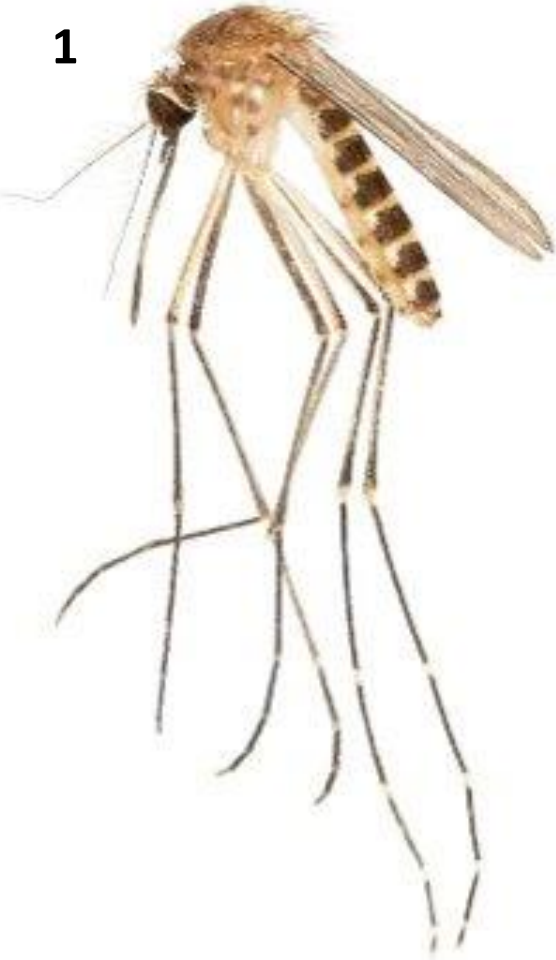


## Pathogen Transmission

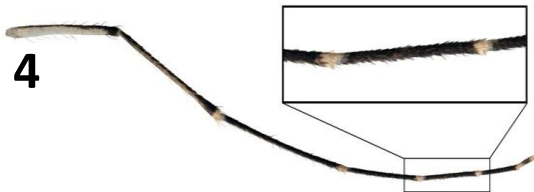
- Babanki virus
- Bunyamwera virus
- Chikungunya virus
- Dengue viruses
- Middleburg virus
- Semliki Forest virus
- Saboya virus
- Wesselsbron virus
- Yellow Fever virus
- Zika virus

# *Culex coronator*

1



1. The adult mosquito is medium sized, drab and brownish



2. The head has dark erected forked scales dorsally. The occiput has narrow golden scales and broad white scales laterally. The proboscis is mostly covered in dark scales, with a ventral median area of pale scales that does not form a complete ring (arrow)

3. The veins of the wings are covered in narrow dark scales

5



4. The hind tarsomeres on the legs are ringed with distinct basal and apical bands

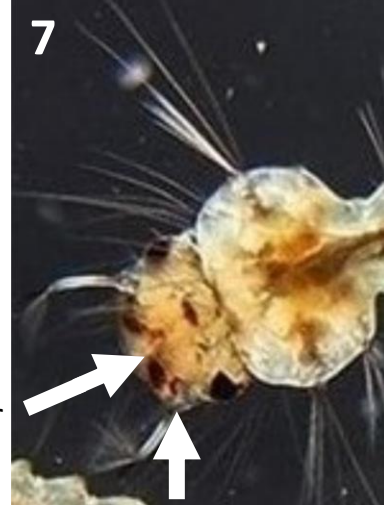
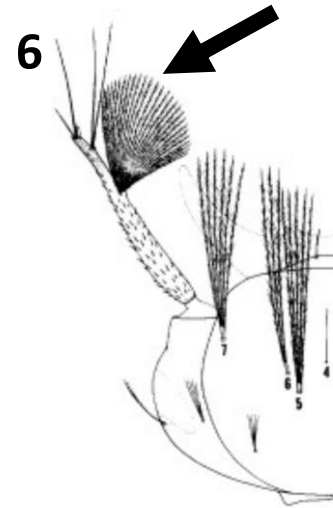
5. Immature four instar larva

6. The antennal turf of the head is located on a constriction near the outer third (arrow), with the shaft spiculated basally

7. The larva head has 4 or 5 upper hairs and 3 or 4 lower hairs

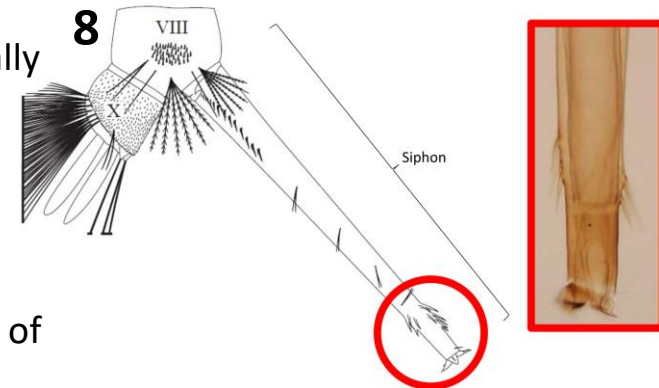
8. The siphon is long and thin with four double siphonal turfs beyond the pecten. Most specimens have a crown of prominent spines.

6



Upper hair

Lower hair



Siphon



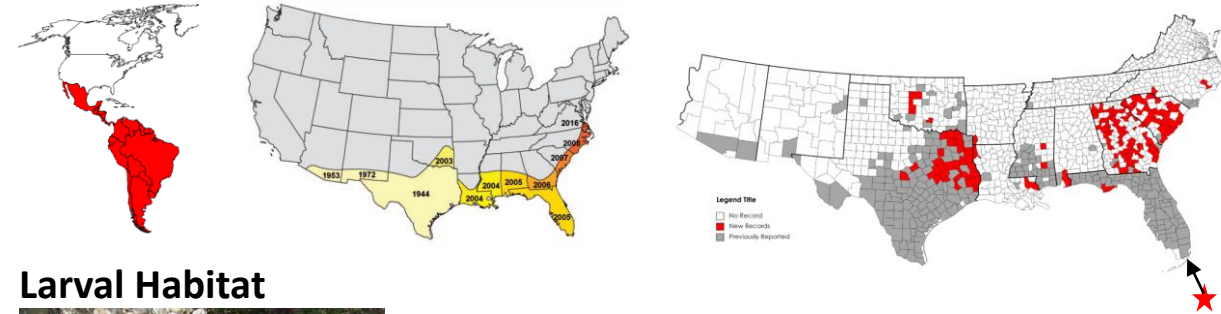




Mosquito BEACONS  
Working Group

# *Culex coronator*

## Native (Red) and Invasive (other colors) Range



## Larval Habitat



- Lay rafts of eggs in diverse natural and artificial microhabitats
- Natural habitats: swales, roadside ditches, animal water troughs, forest ponds, and rock pools
- Artificial water-holding containers: trash cans, car tires

## Dispersal Strategies

- Highly adaptable to artificial container breeding in urban areas
- Eggs and larvae commonly found in used tires
- Car tire transportation is thought to be an important dispersal mechanism

## Collection Methods



- CDC light traps
- Biogents Sentinel traps
- Gravid traps
- Larval surveys

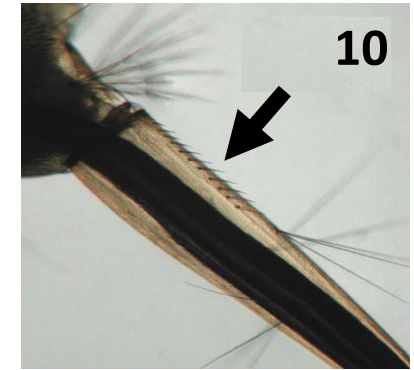
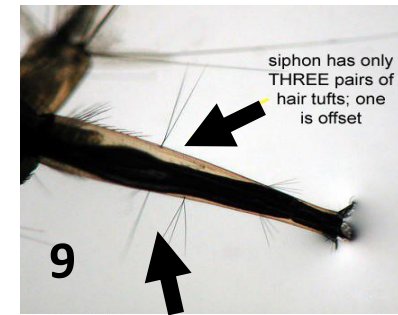
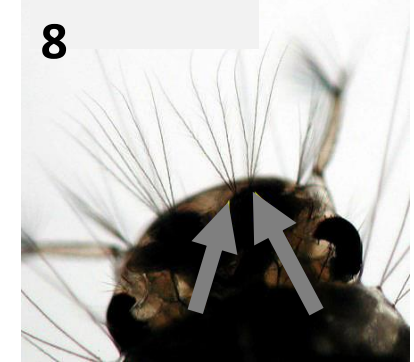
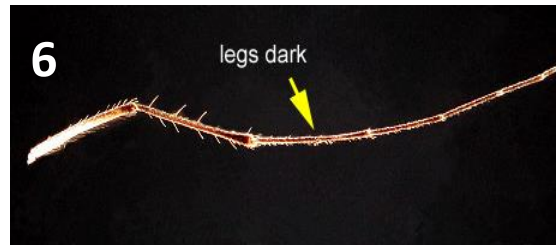
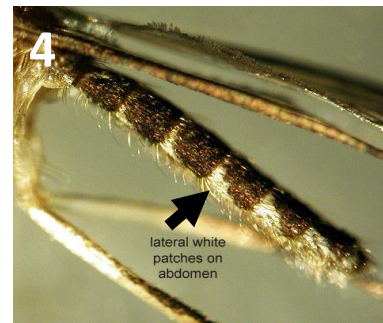
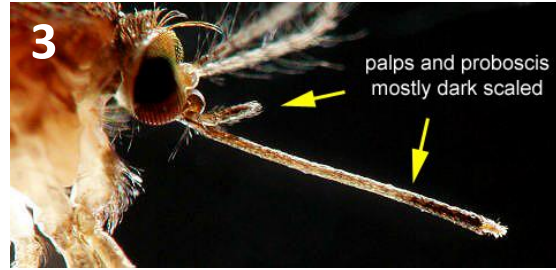
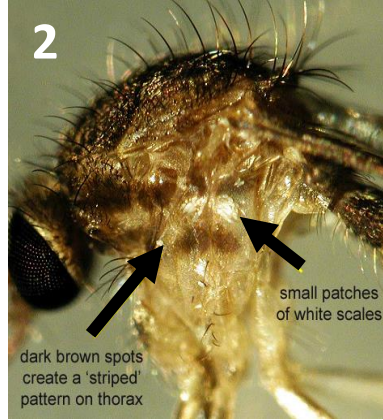
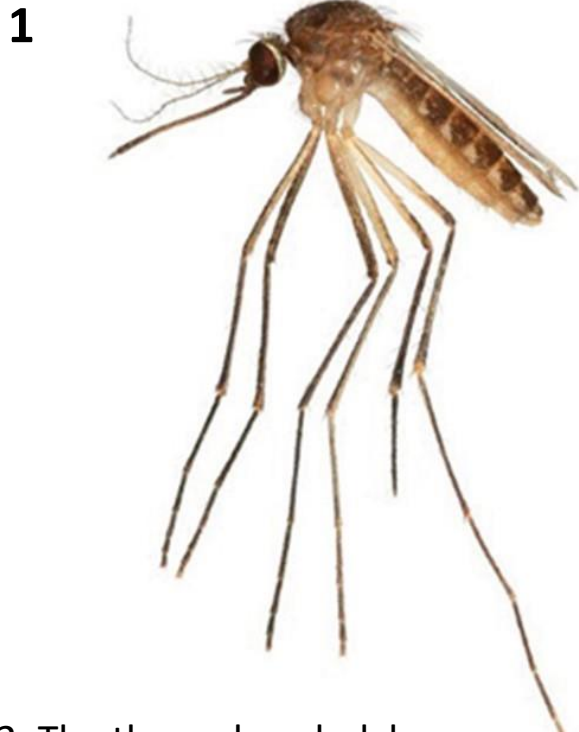
## Host Biting Preference

- Predominantly nocturnal
- Blood feed primary upon large mammals such as white-tailed deer and horses
- Opportunistically feed on birds
- Will also bite humans



## Pathogen Transmission

- St Louis encephalitis virus (Aitken et al. 1964, Turell et al. 2005)
- Venezuelan equine encephalitis virus (Burguete et al. 1973)
- West Nile virus (Mackay 2007, Kelly et al. 2008, Unlu et al. 2010)
- Zika virus (rare, Elizondo-Quiroga et al. 2018)



2. The thorax has dark brown, narrow scales with lateral brown spots that create a striped pattern and lateral small patches of white scales
3. The proboscis has a long pale area on lower surface
4. The abdomen is dark-scaled dorsally with white lateral patches

5. The wing has dark narrow scales
6. Posterior surface of femora and tibiae pale, tarsal segments either dark-scaled or with very narrow white bands
7. Immature larvae

8. The larvae upper and lower head hairs have 3 branches
9. The siphon has 3 pairs of hair tufts; one is offset
10. The pecten teeth is in 1 singly spirally-twisted row



Mosquito BEACONS  
Working Group

# *Culex declarator*

## Native and Invasive Range



## Larval Habitat

- Rural and Suburban habitats
- Tropical and subtropical
- Moist broad leaf forest
- Tree holes
- Close to temporal ponds, swamp and small dams.
- Artificial containers



## Collection Methods

- CDC light traps
- Resting boxes



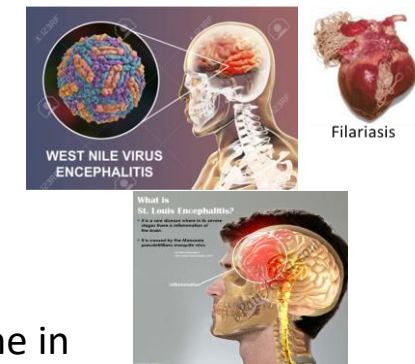
## Host Biting Preference

- Non-human mammals
- Birds
- Less frequently humans



## Pathogen Transmission

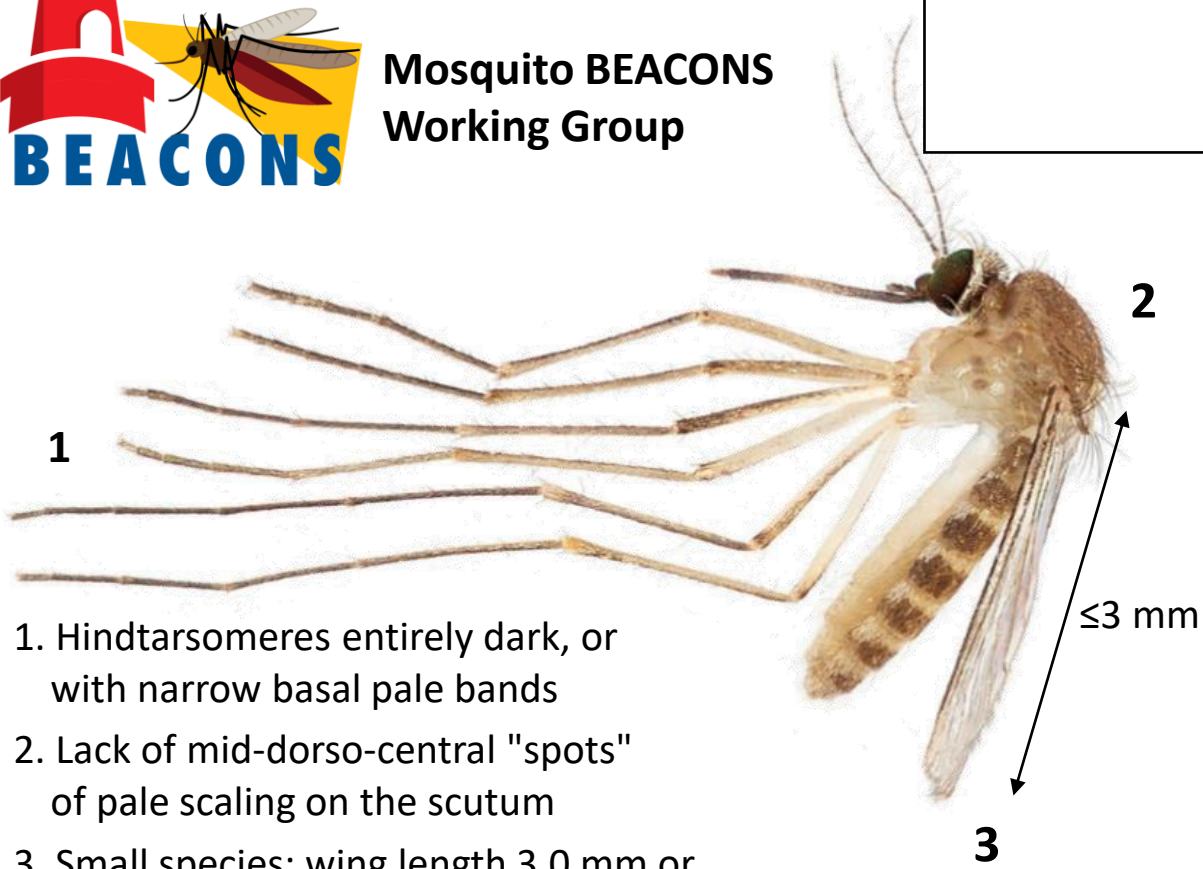
- Saint Louis Encephalitis virus
- West Nile Virus
- Dirofilariasis (dog heartworm)



## Dispersal Strategies

- More research needs to be done in this area

# *Culex interrogator*



1

2

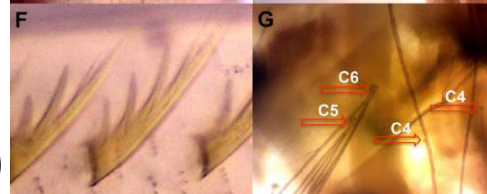
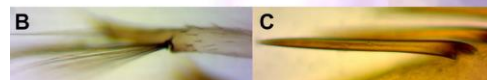
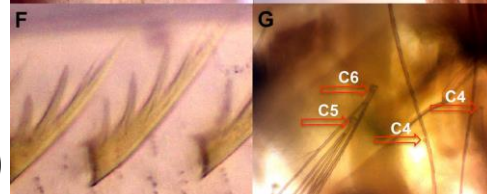
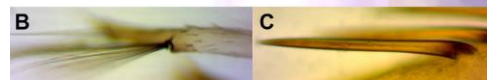
4

3

≤ 3 mm

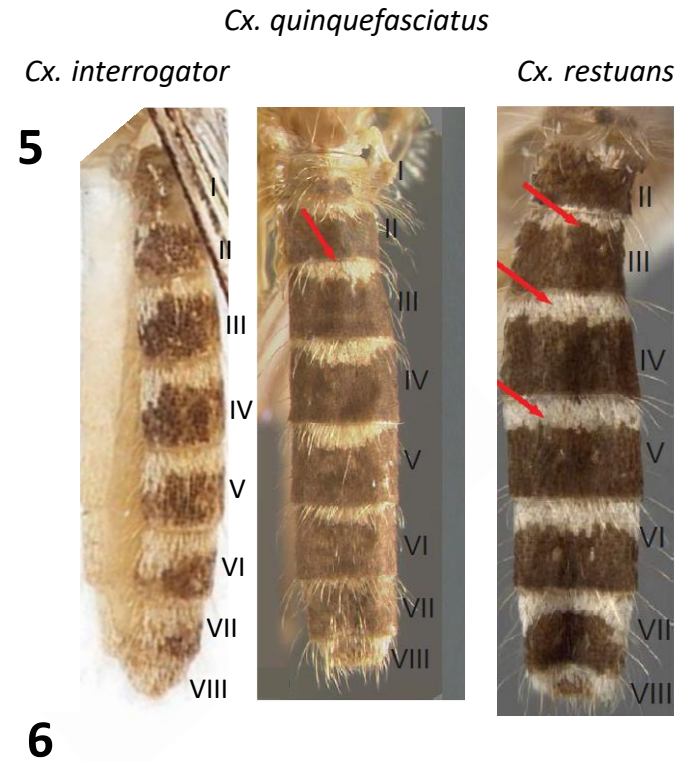


7

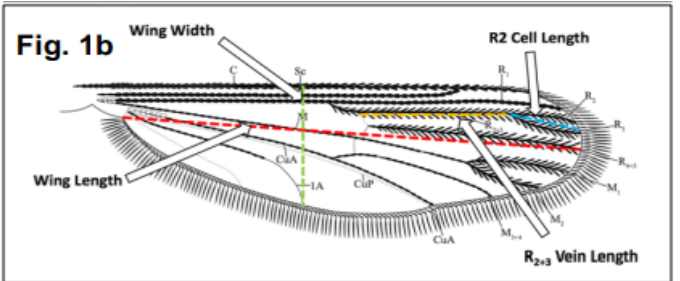


1. Hindtarsomeres entirely dark, or with narrow basal pale bands
2. Lack of mid-dorso-central "spots" of pale scaling on the scutum
3. Small species; wing length 3.0 mm or less.
4. Dark integumental spots on the meskatepisternum and mesepimeron
5. Terga with distinct basal pale bands on segments II–VII. Terga with basal pale bands almost straight posteriorly
6. Cell  $R_2$  usually about 1.2 or less length of vein  $R_{2+3}$

7. *Culex interrogator* larva
  - A. Dorsal view
  - B. Antenna, seta 1–A
  - C. Distal siphonic spine
  - D. Lateral view of the segment VIII, spicules of the pecten
  - E. Lateral view of the siphon
  - F. Basal siphonic spines
  - G. Cephalic setae (C-4, C-5 and C-6)



6



**Fig. 1b** Wing Width, R2 Cell Length, R2+3 Vein Length, Wing Length. **Fig. 1a** Many members of the genus *Culex* remain difficult to morphologically identify, especially when damaged by collection devices such as fan-based traps. The three species we studied in this project include two known West Nile virus (WNV) vectors (*Cx. quinquefasciatus* (Fig.1a) and *Cx. restuans*). The vector status of *Cx. interrogator* remains unknown. We used a combination of wing measurements (Fig.1b) to evaluate potential indices that may be useful for accurately identifying the three species.



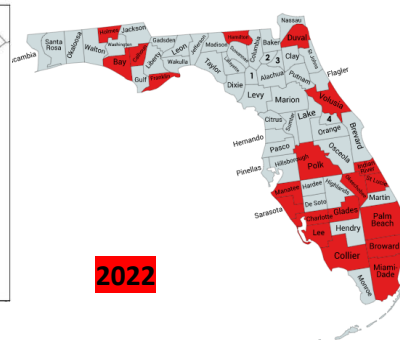
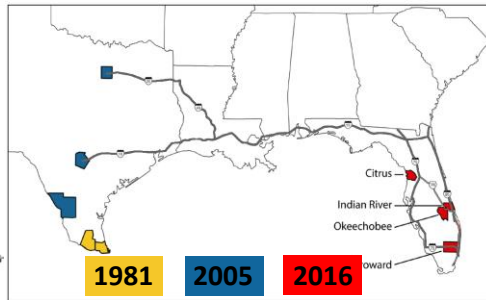
Mosquito BEACONS  
Working Group

# *Culex interrogator*

## Native (Red) and Invasive (green)



## Current Expansion



## Collection Methods



- Significantly lower abundances compared to *Cx. nigripalpus* and *Cx. coronator*
- Adult collections are rare
- Ovitrap
- Gravid traps
- Larval collections

## Larval Habitat



- Permanent and semi-permanent habitats including natural and artificial deposits
- Examples: washtubs, roof gutters, puddles, sewers and manholes

## Dispersal Strategies

- Likely facilitated by movement of mosquito-infested containers (tires, buckets, pots, etc.) along major highway routes

## Host Biting Preference

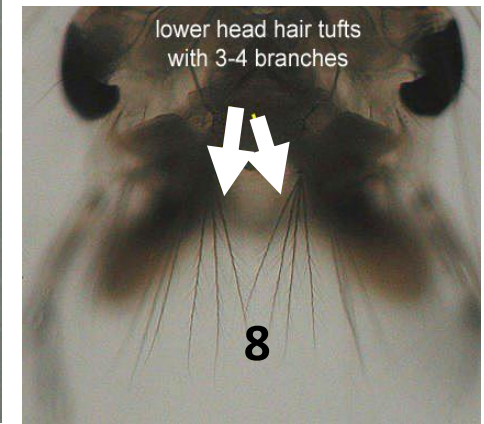
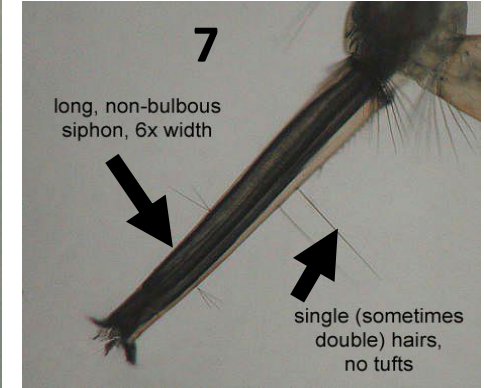
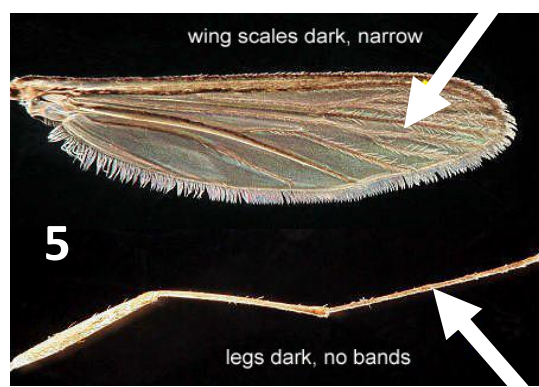
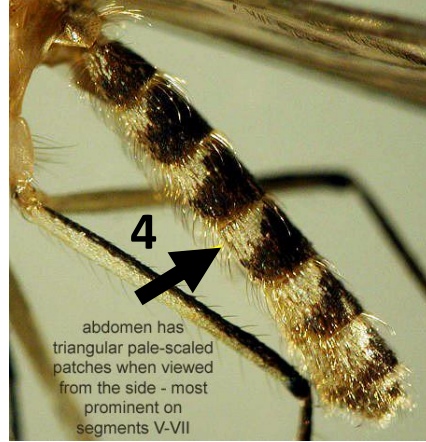
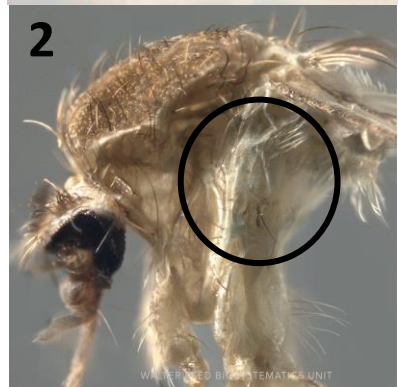
- More surveillance and research is needed in this area
- Collected in dog-baited traps in Mexico
- Association with West Nile virus indicates avian host blood meal



## Pathogen Transmission

- More surveillance and research is needed in this area
- One (1) **West Nile virus** positive mosquito pool identified in Mexico
  - Role in WNV transmission in the United States is unknown
- *Dirofilaria* (dog heartworm parasite) were recovered from one (1) specimen in Mexico
- Insect Specific Virus: *Culex flavivirus*

# *Culex nigripalpus*



- 1. Medium sized dark-brown, dark scaled mosquito species with a rounded abdomen
- 2. No scales present; if present less than 5
- 3. Acrostical seta present; brown scaled scutum

- 4. Triangled shaped pale scaled patches; no bands across tergites
- 5. Wings scaled dark; legs not banded
- 6. Larval Body

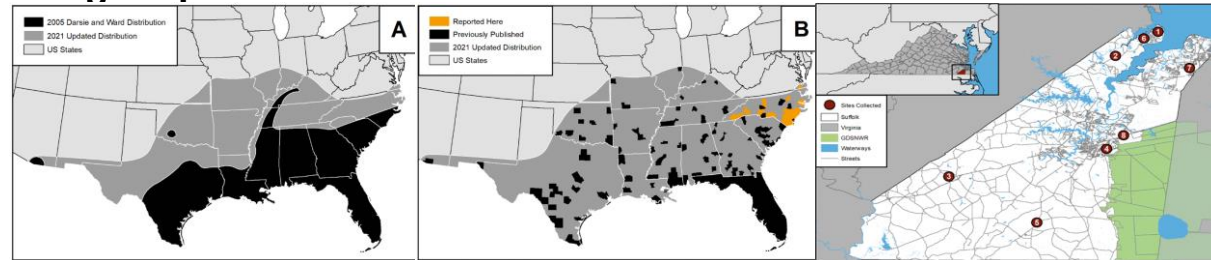
- 7. Long non-bulbous siphon 6X width of base; S-2,3 single sometime double
- 8. 3-C seta with 3 branches



Mosquito BEACONS  
Working Group

# *Culex nigripalpus*

## Range Expansion documented



## Larval Habitat



- Permanent & transient pools
- Freshwater swamps
- Natural & artificial containers
- Effluent ponds
- Brackish water and salt-marsh habitat

## Dispersal Strategies

- 5 km flight range
- Not considered a migratory species
- Increasing temperatures and severe storm systems may be a contributing factor
- Anthropogenic introductions cannot be ignored

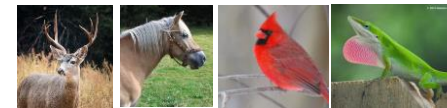
## Collection Methods



- New Jersey Light Traps
- CDC Light Traps
- Canopy Suction Traps
- BG Sentinel Traps
- Aspiration

## Host Biting Preference

- Birds
- Mammals
- Reptiles
- Hypothesized as a major zoonotic transmission contributor



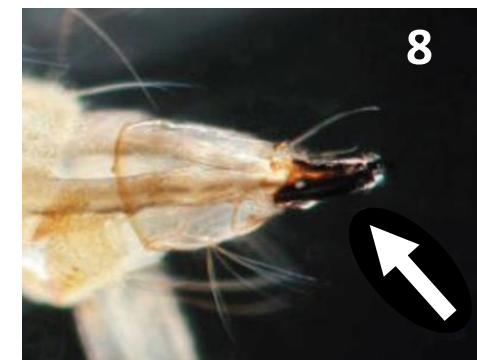
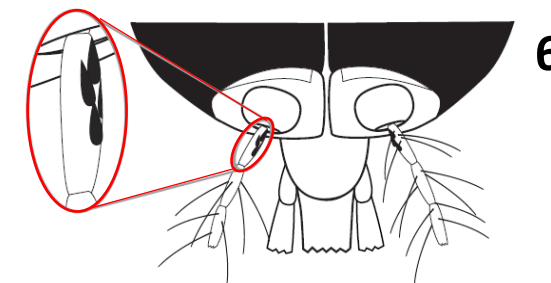
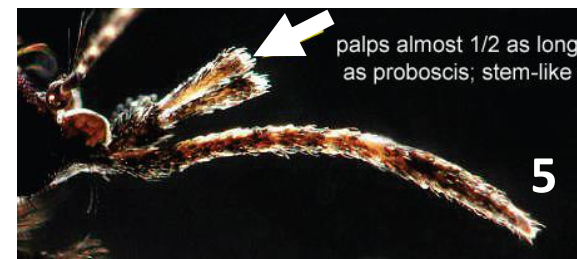
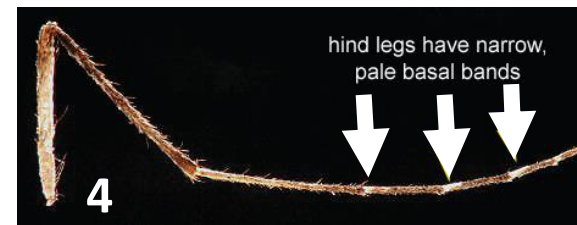
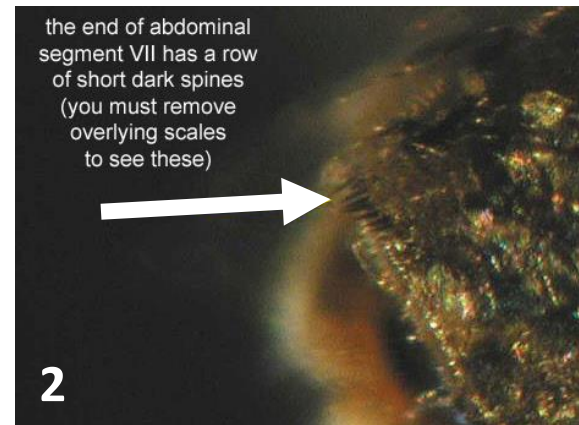
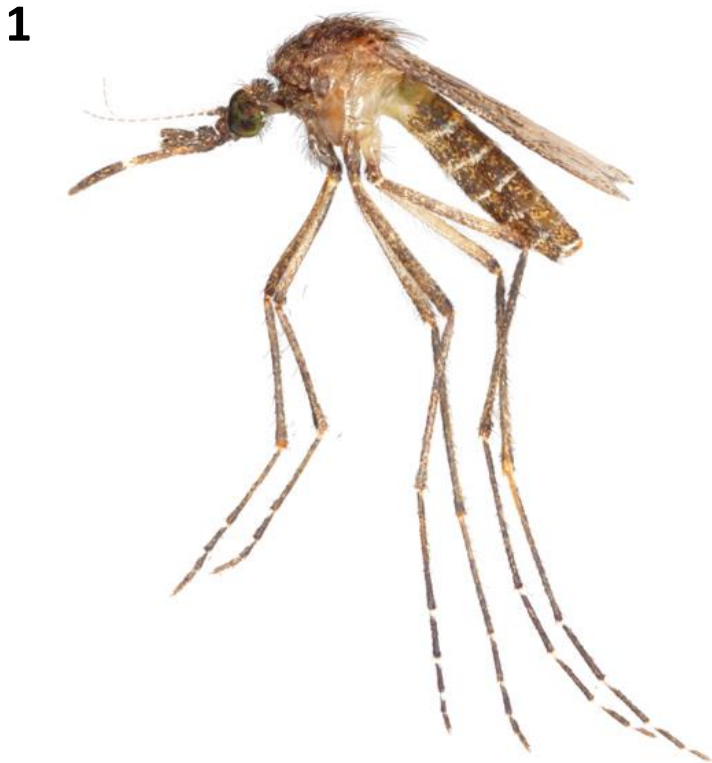
## Pathogen Transmission

- St. Louis encephalitis virus
- West Nile virus



Mosquito BEACONS  
Working Group

# *Mansonia titillans*



1. Medium to large species. Speckled scales throughout. Blunted abdomen; banded proboscis, tarsi and tergites
2. Spiniforms present
3. Pale and dark flat scales present on wing veins

4. Tarsi pale scaled bands below joint
5. Palps almost  $\frac{1}{2}$  length of proboscis
6. Flagellomere I >5 dark scales present

7. Larvae attached to roots of aquatic plant (Photo Credit Michele Cutwa, UF)
8. Modified siphon for piercing roots
9. Antennae seta 1 long and branched (Photo Credit Nathan Burkett-Cadena, UF)

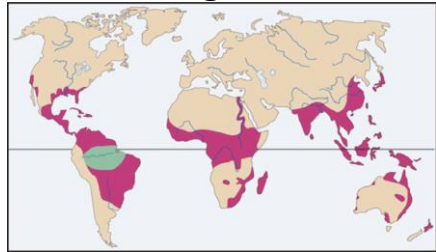




Mosquito BEACONS  
Working Group

# *Mansonia titillans*

## Native Range



- Recently detected in South Carolina
- Last 20 years migration over 15 counties in central and north Florida

## Larval Habitat

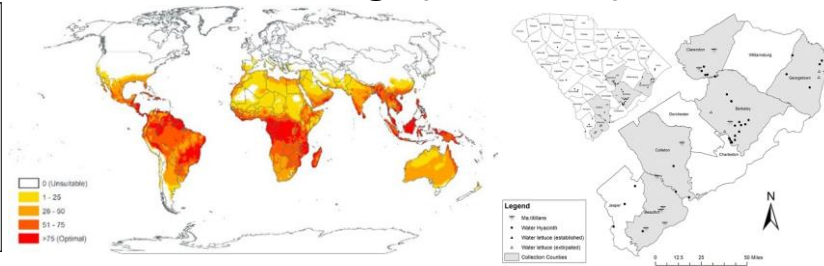


- Permanent bodies of water (lakes, ponds, swamps, etc.) where host plants are found
- Water hyacinth, *Eichhornia crassipes* (top right)
- Water Lettuce, *Pistia Stratiotes* (bottom right)
- Larvae breath using modified larval siphon to puncture plant root and obtain oxygen

## Dispersal Strategies

- Max Flight distance: Known to fly several miles in search of host blood meal and oviposition habitat
- Larvae may be transported in ornamental aquatic plants, explaining relative paucity in SC relative to GA.

## Invasive Range (as of 2021)



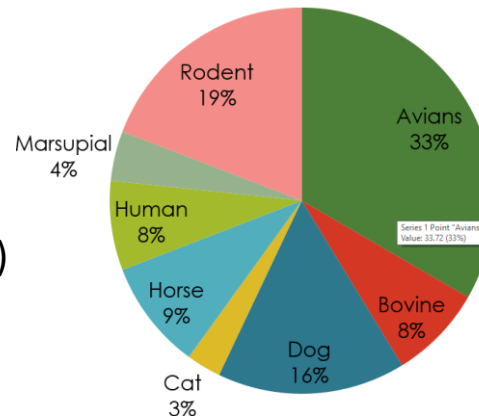
## Collection Methods



- CDC light traps, bait with CO<sub>2</sub>, Octenol, and BG Lure, ideally in synergy
- Emergence traps placed over larval habitat
- Larval collections best done by removing and washing the host plants

## Host Biting Preference

*Mansonia titillans* feeding preference in Brazil



- Generalists, feeding on a combination of mammals, birds, and humans.

## Pathogen Transmission

- Bussuquara virus (BSQV)
- Cabassou virus (CABV)
- St. Louis Encephalitis virus (SLEV)
- Tlacotalpan virus (TLAV)
- Tonate virus (TONV)
- Venezuelan Equine Encephalitis virus (VEEV)
- West Nile virus (WNV)
- *Dirofilaria immitis* (Dog heartworm)

## References

### ***Aedes aegypti***

1. Edman JD, Scott TW, Costero A, Morrison AC, Harrington LC, Clark GG. 1998. *Aedes aegypti* (Diptera: Culicidae) Movement influenced by availability of oviposition sites, *Journal of Medical Entomology* 35:578-583. [doi.org/10.1093/jmedent/35.4.578](https://doi.org/10.1093/jmedent/35.4.578)
2. Etymologia: *Aedes aegypti*. 2016. *Emerging Infectious Diseases* 22:1807. [doi.org/10.3201/eid2210.et2210](https://doi.org/10.3201/eid2210.et2210)
3. Featured creatures [https://entnemdept.ufl.edu/creatures/aquatic/aedes\\_aegypti.htm](https://entnemdept.ufl.edu/creatures/aquatic/aedes_aegypti.htm)
4. Harrington LC, Scott TW, Lerdthusnee K, Coleman RC, Costero A, Clark GG, Jones JJ, Kitthawee S, Kittayapong P, Sithiprasasna R, Edman JD. 2005. Dispersal of the dengue vector *Aedes aegypti* within and between rural communities. *American Journal of Tropical Medicine and Hygiene* 72:209-20. PMID: 15741559.
5. Monaghan, A.J., Eisen, R.J., Eisen, L., McAllister, J., Savage, H.M., Mutebi, J.P. and Johansson, M.A., 2019. Consensus and uncertainty in the geographic range of *Aedes aegypti* and *Aedes albopictus* in the contiguous United States: Multi-model assessment and synthesis. *PLoS computational biology*, 15(10), p.e1007369.
6. Powell JR, Gloria-Soria A, Kotsakiozi P. 2018. Recent history of *Aedes aegypti*: Vector genomics and epidemiology records. *BioScience* 68:854-860.
7. WRBU <https://wrbu.si.edu/vectorspecies/mosquitoes/aegypti>

### ***Aedes albopictus***

1. Bonizzoni M, Gasperi G, Chen X, James AA. 2013. The invasive mosquito species *Aedes albopictus*: current knowledge and future perspectives. *Trends in Parasitology* 2, 460-68.
2. Eritja R, Palmer JRB, Roiz D, Sanpera-Calbet I, Bartumeus F. 2017. Direct Evidence of Adult Dispersal by Car. *Scientific Reports* 7: 14399. <https://www.nature.com/articles/s41598-017-12652-5>
3. Monaghan, A.J., Eisen, R.J., Eisen, L., McAllister, J., Savage, H.M., Mutebi, J.P. and Johansson, M.A., 2019. Consensus and uncertainty in the geographic range of *Aedes aegypti* and *Aedes albopictus* in the contiguous United States: Multi-model assessment and synthesis. *PLoS computational biology*, 15(10), p.e1007369.
4. Giordano BV, Gasparotto A, Liang P, Nelder MP, Russell C, Hunter FF (2020). Discovery of an *Aedes (Stegomyia) albopictus* population and first records of *Aedes (Stegomyia) aegypti* in Canada. *Medical and Veterinary Entomology* 34:10-16.
5. Hofhuis A, Reimerink J, Reusken C, Scholte E-J, de Boer A, Takken W, Koopmans M. 2009. The hidden passenger of lucky bamboo: Do imported *Aedes albopictus* mosquitoes cause dengue virus transmission in the Netherlands? *Vector-borne and Zoonotic Diseases* 9:217-220. DOI: 10.1089/vbz.2008.0071
6. Lacour G. 2016. Eco-physiological mechanisms and adaptive value of egg diapause in the invasive mosquito *Aedes albopictus* (Diptera: Culicidae). [PhD Dissertation, UCL, Belgium]
7. Monaghan, A.J., Eisen, R.J., Eisen, L., McAllister, J., Savage, H.M., Mutebi, J.P. and Johansson, M.A., 2019. Consensus and uncertainty in the geographic range of *Aedes aegypti* and *Aedes albopictus* in the contiguous United States: Multi-model assessment and synthesis. *PLoS computational biology*, 15(10), p.e1007369.

## References

### *Aedes japonicus*

1. <https://www.wrbu.si.edu/vectorspecies/mosquitoes/japonicus>
2. Kaufman MG, Fonseca DM. 2014. Invasion Biology of *Aedes japonicus japonicus* (Diptera: Culicidae). *Annual Review of Entomology* 59(1):31-49. DOI: 10.1146/annurev-ento-011613-162012
3. Lippi, C., Kaufman, P.E. and Buckner, E.A., 2021. Asian Bush Mosquito, Asian Rock Pool Mosquito *Aedes japonicus japonicus* (Theobald, 1901)(Insecta: Diptera: Culicidae). *EDIS*, 2021. [https://entnemdept.ufl.edu/creatures/AQUATIC/aedes\\_japonicus.html](https://entnemdept.ufl.edu/creatures/AQUATIC/aedes_japonicus.html) Accessed November 26, 2022.
4. Riles MT, Smith JP, Burkett-Cadena N, Connelly CR, Morse GW, Byrd BD. First record of *Aedes japonicus* in Florida. *Journal of the American Mosquito Control Association*. 2017 Dec;33(4):340-4.
5. Rogers DC, Goldhammer DS, Garrison I, Cook A. First record of *Aedes japonicus japonicus* (Theobald, 1901)(Diptera: Culicidae) in Kansas. *Zootaxa*. 2021 Sep 23;5040(4):592-600.
6. Sallam, M.F., Ahmed, T., Sylvain-Lear, C., Riegel, C. and Moise, I.K., 2020. Spatial distribution of discarded vehicle tires and their influence on *Aedes albopictus* and *Culex quinquefasciatus* populations, New Orleans, Louisiana. *BioRxiv*. (preprint) <https://www.biorxiv.org/content/10.1101/2020.02.10.942706v1.abstract> Accessed November 26, 2022.
7. Peyton EL, Campbell SR, Candeletti TM, Romanowski M, Crans WJ. *Aedes (Finlaya) japonicus japonicus* (Theobald), a new introduction into the United States. WALTER REED BIOSYSTEMATICS UNIT WASHINGTON DC; 1999 Jan 1.
8. Sames, W.J., Hamik, J., Mann, J.G., Bast, J.D. and Pitts, R.J., 2022. *Aedes japonicus japonicus* in Nebraska and Texas. *Journal of the American Mosquito Control Association*, 38(2), pp.92-95.
9. <https://www.ecdc.europa.eu/en/publications-data/aedes-koreicus-current-known-distribution-march-2021>
10. Photo Credits:  
Matt Flower, some rights reserved (CC BY-NC), <http://www.inaturalist.org/photos/962846>  
Jessie Hirsch, some rights reserved (CC BY), <https://www.flickr.com/photos/jhirsch/2484506458/>

## References

### ***Aedes notoscriptus***

1. Duchemin JB, Mee PT, Lynch SE, Vedururu R, Trinidad L, Paradkar P. 2017. Zika vector transmission risk in temperate Australia: a vector competence study. *Virology* . 14:1-10. <https://doi.org/10.1186/s12985-017-0772-y>
2. Metzger ME, Wekesa JW, Klueh S, Fujioka KK, Saviskas R, Arugay A, et al. 2021. Detection and establishment of *Aedes notoscriptus* (Diptera: Culicidae) mosquitoes in Southern California, United States. *Journal of Medical Entomology*. tjab165. <https://doi.org/10.1093/jme/tjab165>
3. Wilkerson, RC, Linton, YM, & Strickman, D. 2021. Mosquitoes of the World. Volume 1&2. Johns Hopkins University Press, Baltimore, MD.
4. Watson TM, Saul A, Kay BH. 2000. *Aedes notoscriptus* (Diptera: Culicidae) survival and dispersal estimated by mark-release-recapture in Brisbane, Queensland, Australia. *Journal of medical entomology*. 37:380-384. <https://doi.org/10.1093/jmedent/37.3.380>
5. Webb, C., Doggett, S. and Russell, R., 2016. *A guide to mosquitoes of Australia*. Csiro Publishing. pp. 123-124.
6. Photo Credits: iNaturalist contributors andrew\_allen, aaronstevenson, tony\_wills  
[https://www.inaturalist.org/taxa/407524-Aedes-notoscriptus/browse\\_photos](https://www.inaturalist.org/taxa/407524-Aedes-notoscriptus/browse_photos)

### ***Aedes pertinax***

1. Boehmler M, and Hribar LJ. 2022. *Aedes pertinax*, a lost and found new species record for Monroe County, Florida. *Journal of the American Mosquito Control Association* 38(4) <https://doi.org/10.2987/22-7083>
2. Shroyer DA, Harrison BA, Bintz BJ, Wilson MR, Sither CB, Byrd BD. 2015. *Aedes pertinax*, a newly recognized mosquito species in the United States. *Journal of the American Mosquito Control Association*, 31:97-100. DOI:10.2987/14-6447R.1
3. Kara T-J, Reeves L, Lloyd A, Hoel D. 2022. *Aedes pertinax* and *Culex interrogator*: Two mosquito species new to Lee County, Florida. *Journal of the Florida Mosquito Control Association* 69.
4. Kovach, BC, Reeves, LE, Domingo C, L'Heureux SN, Burger GV, Schermerhorn SD and Riles MT. 2022. *Aedes pertinax*, *Anopheles perplexans*, *Culex declarator*, and *Cx. interrogator*: An update of the mosquito species records for Charlotte County, Florida. *Journal of the American Mosquito Control Association*. 38(4):000-000. doi: 10.2987/22-7087. Epub ahead of print. PMID: 36399330.
5. Riles, M.T. and Connelly, C.R., 2020. An update of the mosquito records of Florida counties, USA. *Journal of the American Mosquito Control Association*, 36(2), pp.107-111.
6. Walter Reed Biosystematics Unit (2021). *Aedes atlanticus* species page. Walter Reed Biosystematics Unit Website, <http://wrbu.si.edu/vectorspecies/mosquitoes/atlanticus> (accessed on 23 November 2022).

## References

### ***Aedes scapularis***

1. Reeves LE, Medina J, Miqueli E, Sloyer KE, Petrie W, Vasquez C, Burkett-Cadena ND. 2020. Establishment of *Aedes (Ochlerotatus) scapularis* (Diptera: Culicidae) in Mainland Florida, With Notes on the *Ochlerotatus* Group in the United States. *Journal of Medical Entomology* 58:717–729, <https://doi.org/10.1093/jme/tjaa250>
2. Santos CS, Pie MR, da Rocha TC, Navarro-Silva MA .2019. Molecular identification of blood meals in mosquitoes (Diptera, Culicidae) in urban and forested habitats in southern Brazil. *PLoS ONE* 14: e0212517. <https://doi.org/10.1371/journal.pone.0212517>
3. Campbell LP, Burkett-Cadena ND, Miqueli E, Unlu I, Sloyer KE, Medina J, Vasquez C, Petrie W, Reeves LE. 2021. Potential Distribution of *Aedes (Ochlerotatus) scapularis* (Diptera: Culicidae): A Vector Mosquito New to the Florida Peninsula. *Insects*. <https://doi.org/10.3390/insects12030213>
4. Hribar, L.J. and Cerminara, C., 2021. Rediscovery of *Aedes scapularis* (Diptera: Culicidae) in the Florida Keys. *Florida Entomologist*, 104(2), pp.145-145.

### ***Aedes vittatus***

1. Díez-Fernández A, Martínez-de la Puente J, Ruiz S, Gutiérrez-López R, Soriguer R, Figuerola J. 2018. *Aedes vittatus* in Spain: Current distribution, barcoding characterization and potential role as a vector of human diseases. *Parasites & Vectors* 11:297
2. Pagac BB, Spring AR, Stawicki JR, Dinh TL, Lura T, Kavanaugh MD, Pecor DB, Justi, SA, Linton YM. 2021. Incursion and establishment of the Old World arbovirus vector *Aedes (Fredwardsius) vittatus* (Bigot, 1861) in the Americas. *Acta Tropica* 221:105739.
3. Walter Reed Biosystematics Unit (2022). *Aedes vittatus* species page. Walter Reed Biosystematics Unit Website. [http://wrbu.si.edu/vectorspecies/mosquitoes/ae\\_vittatus](http://wrbu.si.edu/vectorspecies/mosquitoes/ae_vittatus) Accessed on Jan 1, 2022.
4. Wilkerson, RC, Linton, YM, & Strickman, D. 2021. Mosquitoes of the World. Volume 1&2. Johns Hopkins university Press, Baltimore, MD.

## References

### ***Culex coronator***

1. Akaratovic KI, Kiser JP. 2017. First record of *Culex coronator* in Virginia, with notes on its rapid dispersal, trapping methods, and biology. *Journal of the American Mosquito Control Association* 33:225-228.
2. Boehmler, M.B., 2022. *Culex coronator*: A New Species Record For Monroe County, Florida. *Journal of the American Mosquito Control Association*, 38(2): 96-98.
3. Connely. 2008. Is it *Culex tarsalis* or *Culex coronator*? <https://fmel.ifas.ufl.edu/publication/buzz-words/buzz-words-archive/is-it-culex-tarsalis-or-culex-coronator/>
4. Sames, W.J., Mann, J.G., Kelly, R., Evan, C.L., Varnado, W.C., Bosworth, A. B., Noden, B.H., Ramberg, F.B., Riles, M.T., Killingsworth, D, Doyle, M.S., and Pitts, R.J. (2021). Distribution of *Culex coronator* in the USA. *Journal of the American Mosquito Control Association*. 37(1):1-9.

### ***Culex declarator***

1. Barata EA, Chiaravalloti Neto F, Dibo MR, Macoris Mde L, Barbosa AA, Natal D, Barata JM, Andrigueti MT. 2007. Captura de culicídeos em área urbana: avaliação do método das caixas de repouso [Capture of culicids in urban areas: evaluation of the resting box method]. *Rev Saude Publica*. 41:375-382. Portuguese. doi: 10.1590/s0034-89102007000300008.
2. [Culex declarator Dyar & Knab, 1906 \(gbif.org\)](#)
3. Darsie RF Jr, Shroyer DA. 2004. *Culex* (*Culex*) *declarator*, a mosquito species new to Florida. *J Am Mosq Control Assoc*. 20:224-227.
4. Dyar HG, Knab F. 1906. The larvae of Culicidae classified as independent organisms. *J. New York Entomol. Soc.*, 14:169-230.
5. Figueiredo LT. 2000. The Brazilian flaviviruses. *Microbes Infect*. 2:1643-9. doi: 10.1016/s1286-4579(00)01320-4.
6. Florida Medical Entomology Laboratory. *Culex declarator*. [Culex declarator - UF IFAS Florida Medical Entomology Laboratory - University of Florida, Institute of Food and Agricultural Sciences - UF/IFAS \(ufl.edu\)](#).
7. Komar N, Clark GG. West Nile virus activity in Latin America and the Caribbean. *Rev Panam Salud Publica*. 2006 Feb;19(2):112-7. doi: 10.1590/s1020-49892006000200006.
8. Kovach BC, Reeves LE, Domingo C, L'heureux SN, Burger GV, Schermerhorn SD, Riles MT. 2022. Aedes PERTINAX, Anopheles PERPLEXENS, Culex DECLARATOR, AND CX. INTERROGATOR: AN UPDATE OF MOSQUITO SPECIES RECORDS FOR CHARLOTTE COUNTY, FLORIDA. *J Am Mosq Control Assoc*. 2022 Nov 22. doi: 10.2987/22-7087.
9. Laurito M, Hoyos-López R. 2018. First record of *Culex* (*Culex*) *bidens* (Diptera: Culicidae) in Colombia: Taxonomic and epidemiological implications. *Acta Trop*. 188:251-257. doi: 10.1016/j.actatropica.2018.09.010.
10. Reeves LE, Medina J, Miqueli E, Sloyer KE, Petrie W, Vasquez C, Burkett-Cadena ND. 2021. Establishment of *Aedes* (*Ochlerotatus*) *scapularis* (Diptera: Culicidae) in Mainland Florida, With Notes on the *Ochlerotatus* Group in the United States. *J Med Entomol*. 58:717-729. doi: 10.1093/jme/tjaa250. PMID: 33225354.
11. Silva Jdos S, Souto Couri M, de Leão Giupponi AP, Alencar J. 2014. Mosquito fauna of the Guapiaçu Ecological Reserve, Cachoeiras de Macacu, Rio de Janeiro, Brazil, collected under the influence of different color CDC light traps. *J Vector Ecol*. 39:384-94. doi: 10.1111/jvec.12114.

## References

### ***Culex interrogator***

1. Ferreira-de-Freitas L, Thrun NB, Tucker BJ, Melidosian L, 2020. Bartholomay LC. An evaluation of characters for the separation of two *Culex* species (Diptera: Culicidae) based on material from the upper Midwest. *Journal of Insect Science* 20:21. <https://doi.org/10.1093/jisesa/ieaa119>
2. Manrique-Saide P, Bolio-González M, Sauri-Arceo C, Dzib-Florez S, Zapata-Peniche A, *Ochlerotatus taeniorhynchus*: A probable vector of *Dirofilaria immitis* in coastal areas of Yucatan, Mexico. *Journal of Medical Entomology* 45:169–171. <https://doi.org/10.1093/jmedent/45.1.169>
3. Riles, M. T., & Connelly, C. R. (2020). An update of the mosquito records of Florida Counties, USA. *Journal of the American Mosquito Control Association*, 36(2), 107–111.
4. Robinson, P., Sither, C. and Byrd, B.D., 2018. Mosquito wing measurement separate potential West Nile vectors: A morphometric study of three *Culex* species. Poster session presented at the Research and Scholarship Celebration, Western Carolina University. [https://libres.uncg.edu/ir/wcu/f/Robinson\\_RASC18.pdf](https://libres.uncg.edu/ir/wcu/f/Robinson_RASC18.pdf) Accessed November 25, 2022.
5. Shin D, O'Meara GF, Civana A, Shroyer DA, Miqueli E. 2016. *Culex interrogator* (Diptera: Culicidae), a mosquito species new to Florida. *Journal of Vector Ecology* 41: 316-319. <https://doi.org/10.1111/jvec.12230>
6. Sosa, MAR, Rueda, J., Rodríguez, RJP, Bautista, YEV, Tiburcio, JCD, Fimia-Duarte, R. and Alarcón-Elbal, PM, 2020. First record of *Culex interrogator* (Diptera: Culicidae) for Hispaniola and update of the list of mosquitoes from Jarabacoa, Dominican Republic. *Novitates Caribaea* , (16), pp.110-121.
7. Ulloa A, Ferguson HH, Méndez-Sánchez JD, Danis-Lozano R, Casas-Martínez M, Bond JG, et al. . 2009. West Nile virus activity in mosquitoes and domestic animals in Chiapas, México. *Vector-borne and Zoonotic Diseases* 9:555-560. DOI: 10.1089/vbz.2008.0087

## References

### ***Culex nigripalpus***

1. Akaratovic KI, Kiser JP, Whitt PB, Harrison RL, Harrison BA. 2021. *Culex nigripalpus* Distribution Expansion: First Record in Virginia, New County Records in North Carolina, and Revised United States Map. *Journal of the American Mosquito Control Association* 37:188-97. <https://doi.org/10.2987/21-7025>
2. Hancock, C. and Camp, J.V., 2022. Habitat-Specific Host Selection Patterns of *Culex quinquefasciatus* and *Culex nigripalpus* in Florida. *Journal of the American Mosquito Control Association*, 38(2), pp.83-91.

### ***Mansonia titillans***

1. Santos CS, Pie MR, da Rocha TC, Navarro-Silva MA .2019. Molecular identification of blood meals in mosquitoes (Diptera, Culicidae) in urban and forested habitats in southern Brazil. *PLoS ONE* 14: e0212517. <https://doi.org/10.1371/journal.pone.0212517>
2. Cartner RL, Evans CL, Harrison BA, Hager EJ. 2018. New county records demonstrating a northern expansion of *Mansonia titillans* in South Carolina, USA. *Journal of the American Mosquito Control Association*, 34:134-137. <https://doi.org/10.2987/18-6733.1>
3. Moulis RA, Peaty LF, Heusel JL, Lewandowski Jr HB, Harrison BA, Kelly R, Hager EJ. 2015. *Mansonia titillans*: new resident species or infrequent visitor in Chatham County, Georgia, and Beaufort County, South Carolina, USA. *Journal of the American Mosquito Control Association* 31:167-71. <https://doi.org/10.2987/15-6491R>
4. Morris C, Larson V, Lounibos L. 1991. Measuring mosquito dispersal for control programs. *Journal of the American Mosquito Control Association* 7:608–615.
5. Kriticos DJ, Brunel S. 2016. Assessing and Managing the Current and Future Pest Risk from Water Hyacinth, (*Eichhornia crassipes*), an Invasive Aquatic Plant Threatening the Environment and Water Security. *PLoS ONE* 11: e0120054. <https://doi.org/10.1371/journal.pone.0120054>
6. Harrison, B.A., Byrd, B.D., Sither, C.B. and Whitt, P.B., 2016. *The mosquitoes of the Mid-Atlantic region: an identification guide* (p. 69). Cullowhee, NC: Western Carolina University.
7. Rojas-Araya D, Mathias D, Burkett-Cadena N. 2021. A mosquito *Mansonia titillans* (Walker) (Insecta: Diptera: Culicidae: Culicinae: Mansoniini). <https://edis.ifas.ufl.edu/pdf/IN/IN131400.pdf>